Engineered human IL-2/IL-2Rβ orthogonal pairs selectively enhance anti-GPC3 CAR T cells to drive complete responses in solid epithelial tumor models

Paul-Joseph Aspuria, Marie Semana, Mahalaksmi Ramadass, Navneet Ratti, Sandro Vivona, Romina Riener, Michele Bauer, Mohammed Ali, Deepti Rokkam, Rob A. Kastelein, Patrick J. Lupardus, and Martin Oft Synthekine, Menlo Park, CA



Abstract

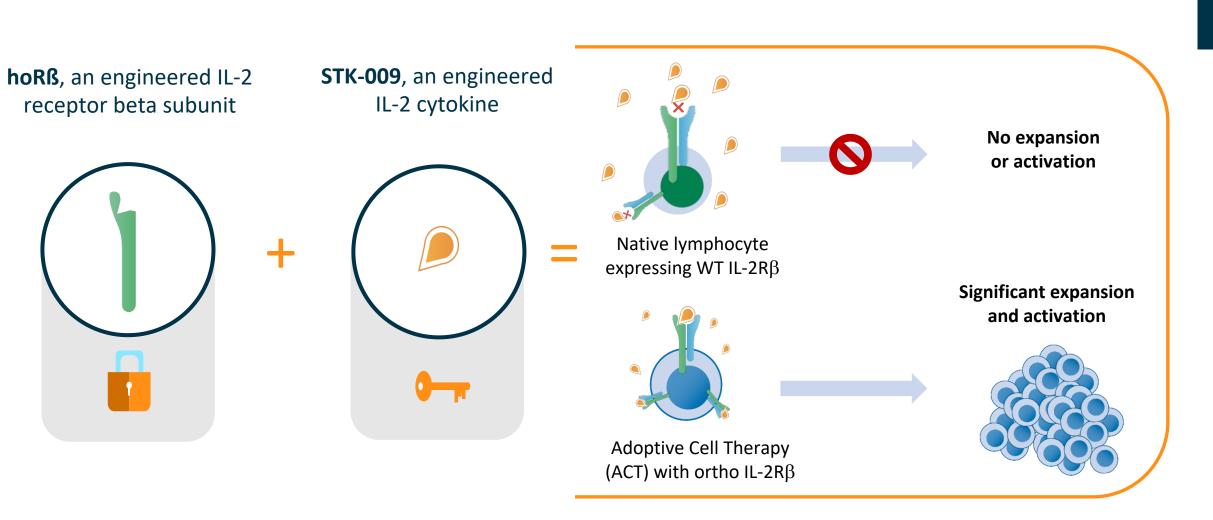
CAR T cell therapy has demonstrated clinical efficacy against relapsed and refractory hematological malignancies. However, prominent barriers including poor T cell effector function, lack of proliferation, and limited CAR T cell persistence have prevented CAR T cell therapies from reaching their full curative potential, especially in solid tumors. Interleukin-2 (IL-2) is a potent stimulator of T cell proliferation, survival, and cytotoxic function, thereby making it an attractive cytokine to support CAR T cell therapy. However, therapeutic use of IL-2 is limited by systemic toxicity due its promiscuous activation of undesired immune cell populations, including non-tumor reactive T cells and NK cells.

To facilitate selective ex vivo and in vivo expansion of engineered T cells we have developed a human orthogonal (ortho) ligand/receptor system consisting of a IL-2 mutein (STK-009) that does not significantly stimulate cells expressing wild type IL-2 receptor and a mutated IL-2 Receptor Beta (orthoIL-2R β) that responds to STK-009 but not wild type IL-2. This system enables *in vivo* IL-2 signaling in engineered cells that express the orthoIL-2R β while avoiding stimulation of native T cells and NK cells. Previously, we demonstrated the ability of the STK-009/orthoIL-2R β receptor pair to selectively enhance the anti-tumor efficacy of orthoIL-2R β (hoRb) expressing CD19 CAR T cells (SYNCAR-001) in preclinical lymphoma mouse models¹. We also demonstrated that STK-009 is selective for the orthoIL-2R β expressing cells and therefore in a non-human primate model does not stimulate native T or NK cells¹.

Here, we demonstrate the ability of the STK-009/hoRb system to enhance the anti-tumor activity and persistence of anti-glypican 3 (GPC3) CAR T cells. GPC3 overexpression is associated with various malignancies such as hepatocellular carcinoma (HCC), pediatric sarcomas, and non small cell lung carcinoma (NSCLC). Clinical trials of GPC3 CAR T therapy are ongoing, but early data suggests a need to boost CAR T cell function and persistence to achieve significant clinical responses. We incorporated the hoRb downstream of a second generation anti-GPC3 CAR via a T2A cleavage peptide (SYNCAR-002) to allow for bicistronic expression from one lentiviral construct. In vivo, STK-009 administration enhanced the anti-tumor efficacy of SYNCAR-002 in highly aggressive subcutaneous (HEPG2) and intraperitoneal (Huh-7) HCC models. STK-009 treatment resulted in significant expansion of SYNCAR-002 in the peripheral blood and drove infiltration of SYNCAR-002 into tumors. STK-009 treatment also induced intratumoral granzyme B and IFN- γ production by SYNCAR-002 indicating activation of effector T cell function.

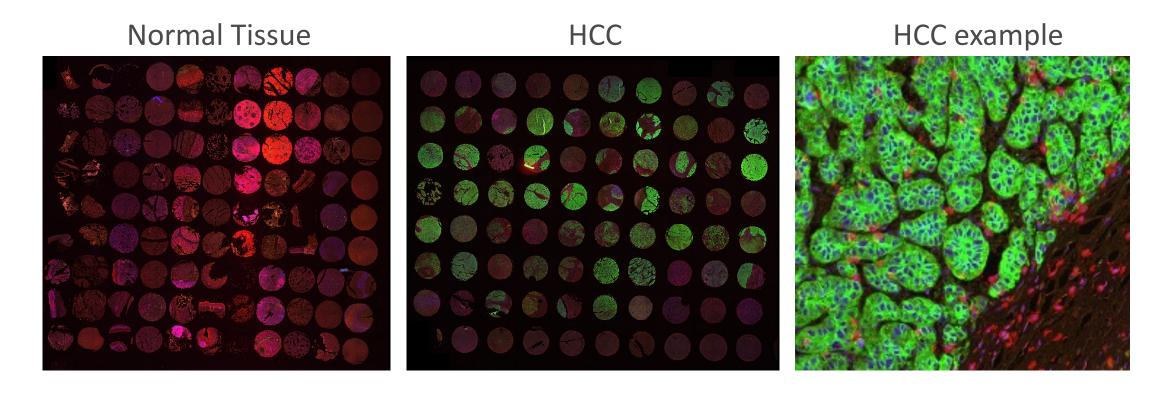
These findings validate that the orthogonal IL-2 platform has the potential to improve the efficacy and durability of CAR T therapy for solid tumor targets such as GPC3 by selectively expanding CAR-T cells in vivo, driving CAR-T cells into the tumor, and activating CAR-T cells in the tumor microenvironment.

Orthogonal Cytokine + Cell Therapy: A Lock and Key System to Stimulate ACTs Selectively In Vivo



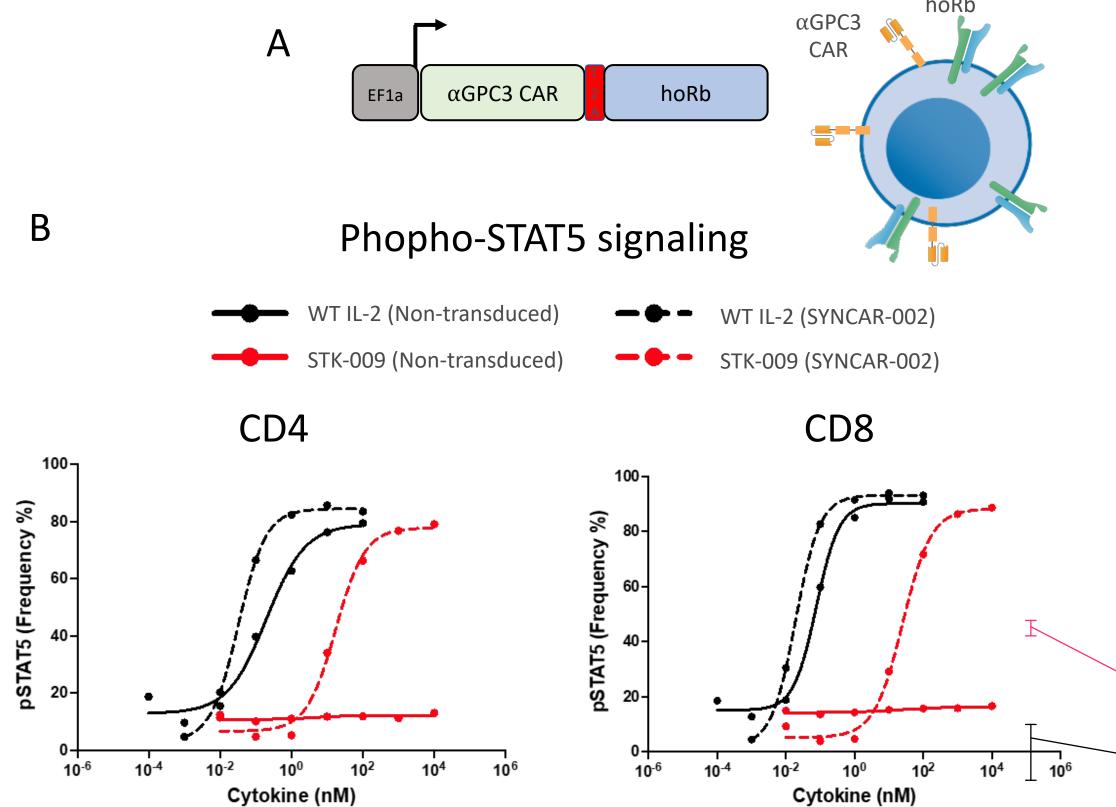
OrthoIL-2Rβ receptors (hoRb) exhibit significant preference for their cognate ligand, STK-009, a pegylated orthogonal IL-2. Therefore, engineered T cells expressing the *ortho* receptor will respond selectively to STK-009, thereby allowing specific expansion and enhancement of engineered T cell activity.

GPC3 is overexpressed in HCC and largely devoid in normal adult tissues



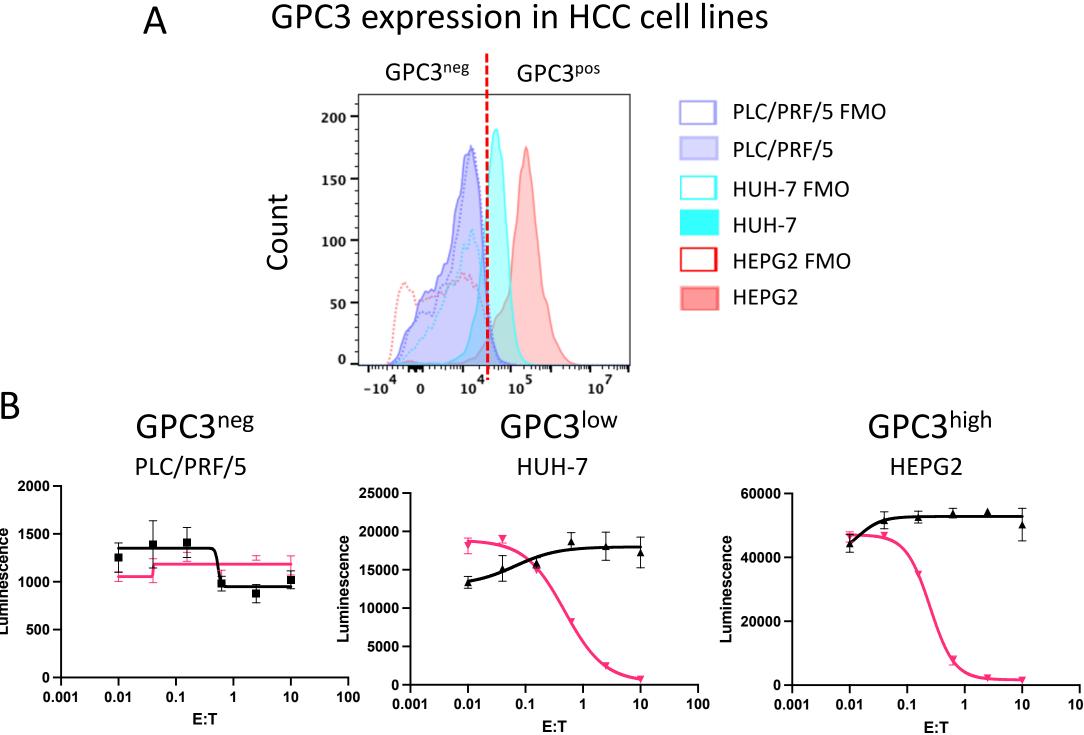
Immunofluorescent staining of GPC3 (green) and CD3 T cells (red). Normal tissue microarray consisting of 90 cores representative of adult healthy tissues. 2/3 of HCC cores stained positive for membranous GPC3 expression. Zoomed in example of a GPC3 positive core.

SYNCAR-002: Anti-GPC3 CAR T cells co-expressing hoRb that specifically respond to STK-009 and maintain response to WT IL-2



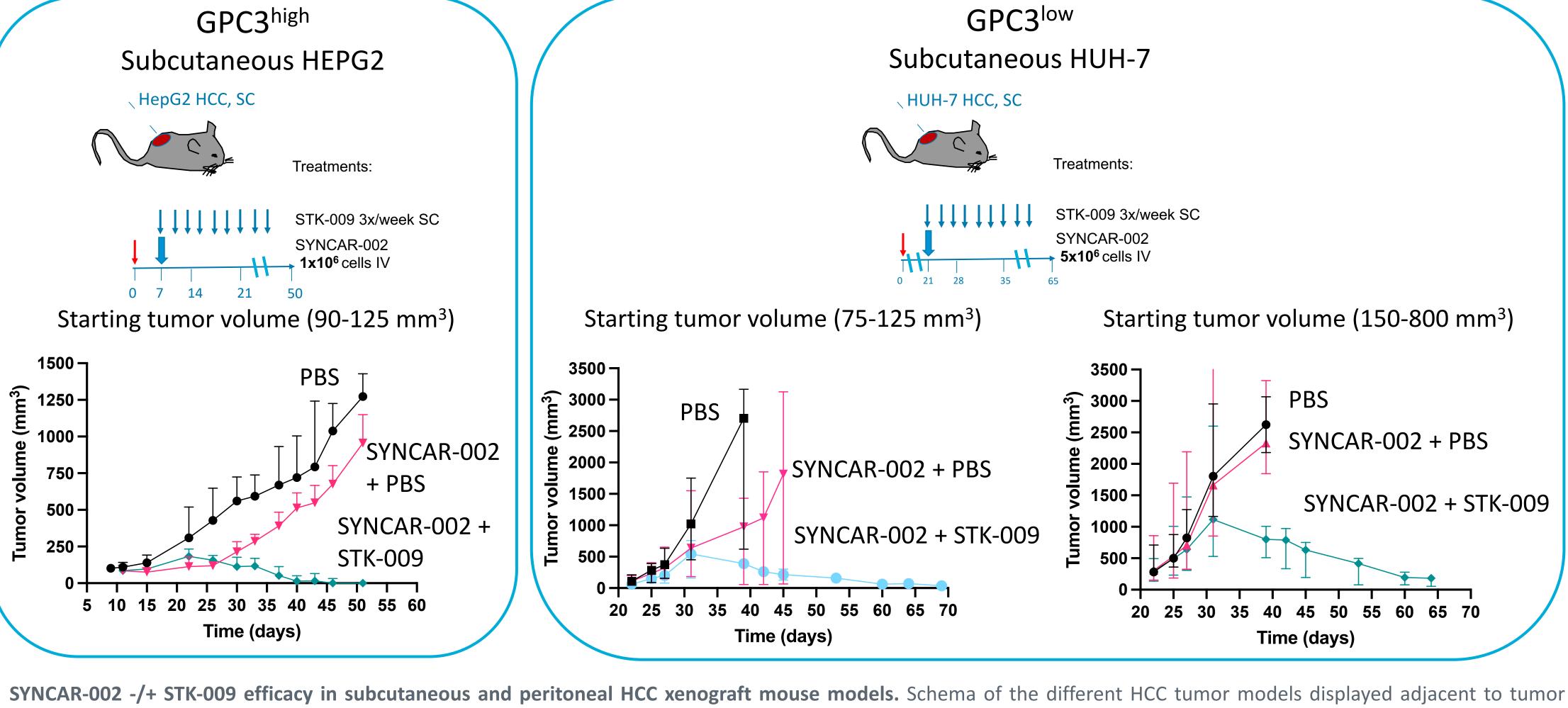
SYNCAR-002 architecture and specific response to STK-009. (A) Lentiviral construct containing the anti-GPC3 CAR, cleavage peptide T2A, and human orthoIL-2R β (hoRb) that immediately follows the CAR construct are expressed as a single mRNA. Expression is regulated via an EF1 α promoter. (B) Phospho-STAT5 signaling assay. Non-transduced and SYNCAR-002 cells were treated with either WT IL-2 or STK-009 for 20 min and processed for pSTAT5 analysis via flow cytometry. pSTAT5 frequency of CD4 and CD8 cells is displayed.

SYNCAR-002 T cells specifically kill GPC3 expressing HCC cell lines

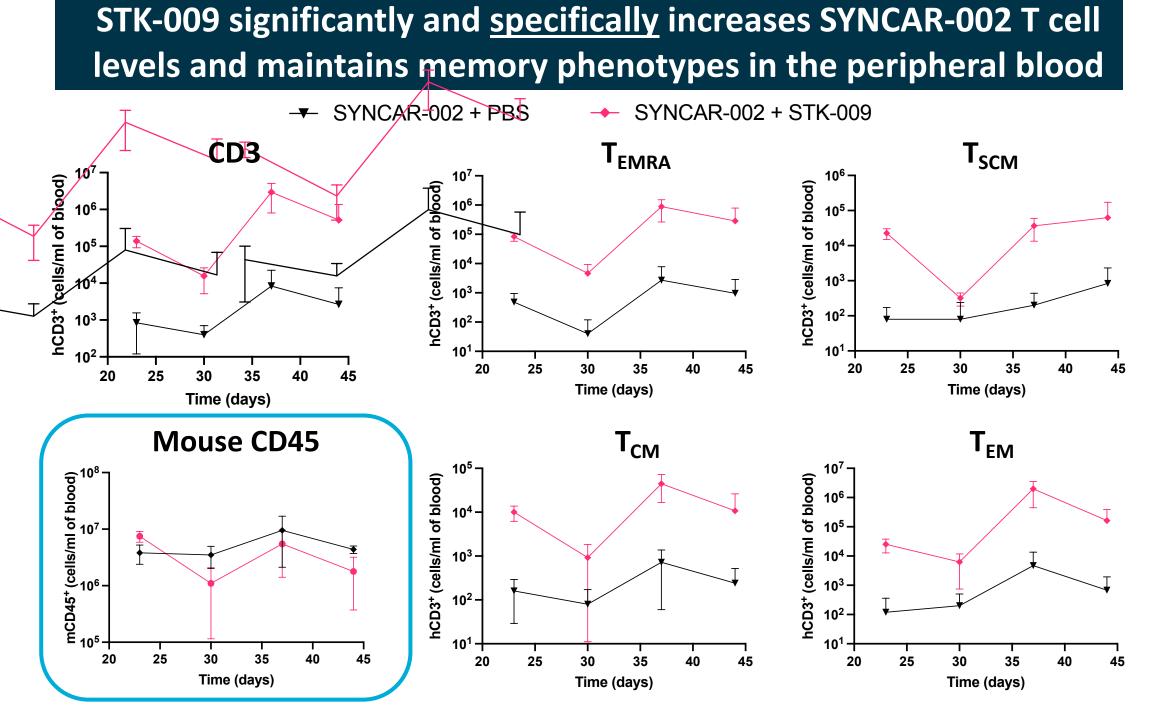


GPC3 expression on HCC cell lines and SYNCAR-002 GPC3-specific cytotoxicity. (A) HCC cell lines stained with anti-GPC3 ab to assess GPC3 expression. Fluorescence minus one (FMO) included as a negative control. (B) SYNCAR-002 (pink) and non-transduced (black) T cells co-cultured with luciferase-expressing HCC lines at indicated effector to target (E:T) ratios. Luminescence equates to cell viability.

STK-009 administration enhances the anti-tumor efficacy of a <u>suboptimal</u> dose of SYNCAR-002 T cells in subcutaneous and intraperitoneal GPC3 expressing HCC mouse models



volume or tumor burden graphs. Subcutaneous models were treated by SYNCAR-002 T cells with or without subcutaneous injection of STK-009 at indicated starting volume. Median tumor volume and tumor burden is displayed.

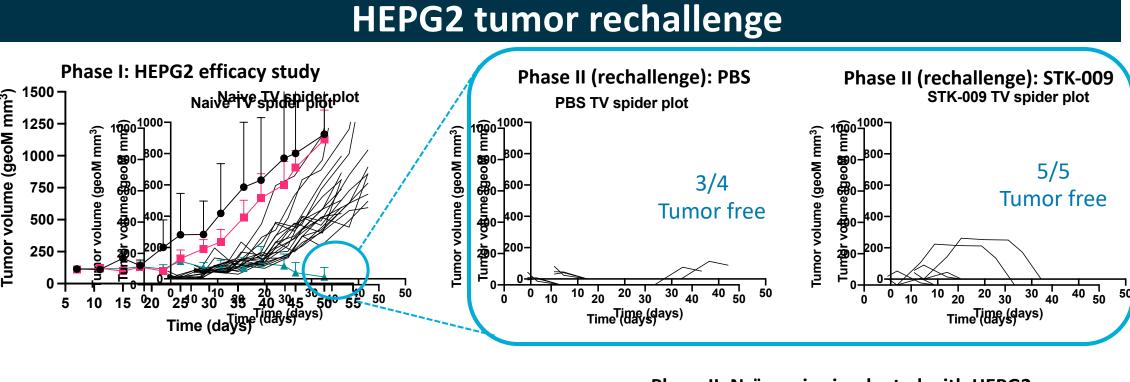


Assessment of peripheral SYNCAR-002 T cell levels and memory phenotypes. From a subcutaneous HEPG2 efficacy study, weekly bleeds were taken 2 weeks post SYNCAR-002 transfer. Cells were stained for mouse CD45, human CD3, CCR7, and CD45RA.

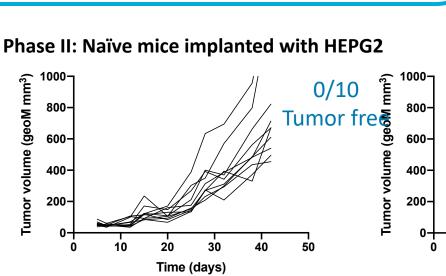
STK-009 significantly increases intratumoral SYNCAR-002 T cell levels and activity SYNCAR-002 + PBS SYNCAR-002 + STK-009 CD3 CD8 Granzyme B Nuclei

Assessment of intratumoral SYNCAR-002 T cell levels and activation. For immunofluorescence, HEPG2 tumors were taken down 14 days post-SYNCAR-002 T cell transfer. Top panel: hCD3 (red) and GPC3 (green). Bottom panel: hCD8 (red), hGranzyme B (green), and nuclei (blue).

Mice previously cured by SYNCAR-002 + STK-009 withstand



subcutaneous HEPG2 tumors that received SYNCAR-002 at Day 7 and subsequent doses of STK-009 were cured by Day 50. HEPG2 + Matrigel were implanted into these mice at Day 55 and into naïve mice. Previously cured mice were split into two groups receiving either PBS (n=4) or STK-009 2x/week (n=5).



CONCLUSION

Adoptive T cell therapies, including CAR T cells, are increasingly recognized as important modalities in the treatment of cancer. However, CAR Ts and other ACTs suffer from several challenges that limit their effectiveness and utilization, including poor expansion and short persistence *in vivo*, diminished activity in the tumor microenvironment, and toxicities related to high initial CAR-T doses and uncontrolled expansion. We have designed the STK-009/orthoIL2R β to address these challenges with CAR-Ts. SYNCAR-002, a GPC3 CAR-T powered by STK-009, demonstrates:

- Improved anti-tumor efficacy in HCC solid tumor xenograft models
- Increased levels and persistence of CAR T cells in vivo
- Increased activation of intratumoral CAR T cells to overcome a hostile tumor microenvironment

Therefore, the STK-009/SYNCAR platform has the potential to overcome clinically relevant hurdles in cell therapy especially in solid epithelial tumor indications.